



NIZAM COLLEGE

Autonomous
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A Constituent College of Osmania University



Two days workshop on latest technologies in Rapid Microbial testing 22nd and 23rd February 2023

Program

22.02.23 Wednesday

10-11am Inaugural

11-11.30am Tea break

11.30am to 1 pm Chromagar based bacterial testing.

1-2 pm Lunch

2-4.30 pm Experiments on Chromagar based bacterial testing.

23.02.23 Thursday

10-11am Taking results of Chromagar based bacterial testing.

11-12am Advances in ELISA for detection of Microbes,
Mycotoxins, Allergens, & Antibiotic etc.

12-1pm Extraction for ELISA

1-2 Pm Lunch

2-4 ELISA of Microbes Mycotoxins, Allergens, & Antibiotic

4Pm Valedictory Function

Contact

Resource Person

Dr. Navneeth Saxena

R-Biopharma Neugen Pvt Ltd

Dr Chand Pasha

hodmicrobiology@nizamcollege.ac.in

Phone: 9441031626

Chief Patron

Prof D.Ravinder,
Vice chancellor, OU

Patron

Prof B.Bhima
Principal Nizam College

Organizing Secretary

Dr Chand Pasha
HoD Microbiology, NC

Organizing Committee

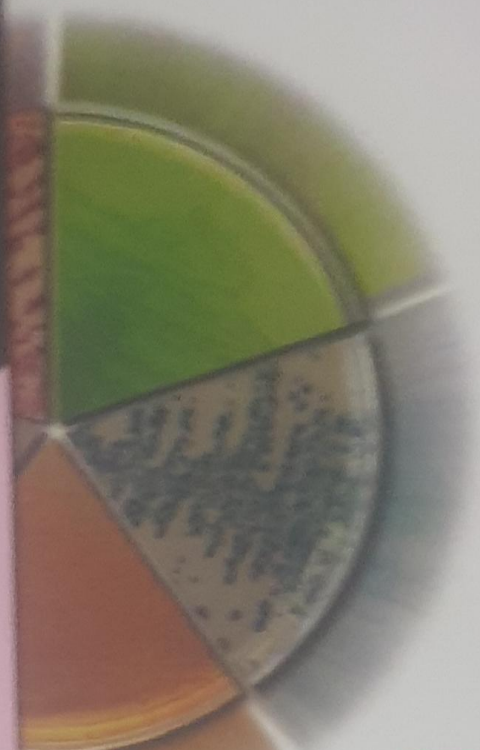
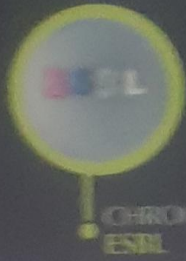
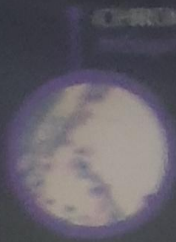
Dr Sandepta Burgula
Dr Hameeda Bee
Dr Shanti Kumari
Dr SambaShiva
Dr Anju Nitin Rajan

Only the Microbiology faculty members and students 15 each will be selected as first come first serve basis for free workshop. Apply through the following link

<https://docs.google.com/forms/d/e/1FAIpQLSfTwuRnlCJW-PsdxWWdvopsSdOJTveoC4xEDqV8yxCVvEG14Q/viewform?vc=0&c=0&w=1&flr=0>

Organized by the Department of Microbiology, Nizam College, Osmania University

Enumeration Identification Organisms in urine



CHROMagar™ solutions
for Drug resistant bacteria
Detection & Surveillance

CHROMagar™
The Chromogenic Media Pioneer

22-02-2023 Wednesday
E. coli,

Staphylococcus aureus, Klebsiella pneumoniae, Pseudomonas aeruginosa and Enterococcus.

- Pre-poured or plated media
- Isolation from urine within 18 - 24 hours
- Rapid, visual enumeration
- Chromogenic and conventional substrates
- Specific identification or orientation

URICHROM II

SALMONELLA

25 g Sample (Tomato)

[Primary Enrichment]

↓
225 ml Broth

**sample preparation for
Salmonella detection in Tomato**

↓
Incubate for 16 ± 24

35 - 37°C [Secondary Enrichment]

↓
Ramba Quick Broth - 42°C

- 7 hrs

0.1 ml

U 10ml

Results

Klebsiella

Salmonella

ZEHRA
22/2/23

70.7



SAMPLE PREPERATION FOR NITROFURON IN SHRIMPS

(SEM)

- Mix 1 g of the homogenized sample Add 4 ml distilled water, 0.5 ml 1 M HCl+(Add spike for validation)
- Add 100 µl 10 mM 2-Nitrobenzoic aldehyde (in DMSO) by shaking vigorously
- Incubate at 50 °C for 3 hours or at 37 °C overnight (approx.16 h)
- Add 5 ml 0.1 M K₂HPO₄+ 0.4 ml 1 M NaOH+ 10 ml ethyl acetate,
- Shake Vigorously for 30 sec.
- Centrifuge 10 min / 3000rpm / at room temperature
- Transfer 5 ml of the ethyl acetate layer (upper layer) into a new vial and Evaporate
- Dissolve the residue in 1 ml n-hexane (or n-heptane) and mix properly with 1ml sample buffer.
- Centrifuge 10 min / 3000 g / at room temperature.
- Use 50 µl of the lower, aqueous phase per well in the assay. (For SEM)

Elisa Test Procedure:-

ELISA

- Insert a sufficient number of wells into the microwell holder for all standards and samples to be run. Record standard and sample positions.
- Pipet 50 µl of standard or prepared sample into separate wells. Use a new pipette tip for each standard or sample.
- Add 50 µl of conjugate to each well.
- Mix gently by shaking the plate manually.
- incubate for 30 min (+/- 1) at room temperature
- Pour the liquid out of the wells and tap the microwell holder vigorously upside down against absorbent paper (three times in a row) to ensure complete removal of liquid from the wells. Fill all the wells with 250 µl wash buffer. Empty the wells again. Repeat two more times.
- Add 100 µl of substrate/chromogen to each well.
- Mix gently by shaking the plate manually and incubate for 15 min (+/- 1) at room temperature.
- Add 100 µl of stop solution to each well.
- Mix gently by shaking the plate manually and measure the absorbance at 450 nm. Read within 15 minutes after addition of stop solution.

23-02-2023 Thursday

RIDASCREEN Aflatoxin B1 30/15

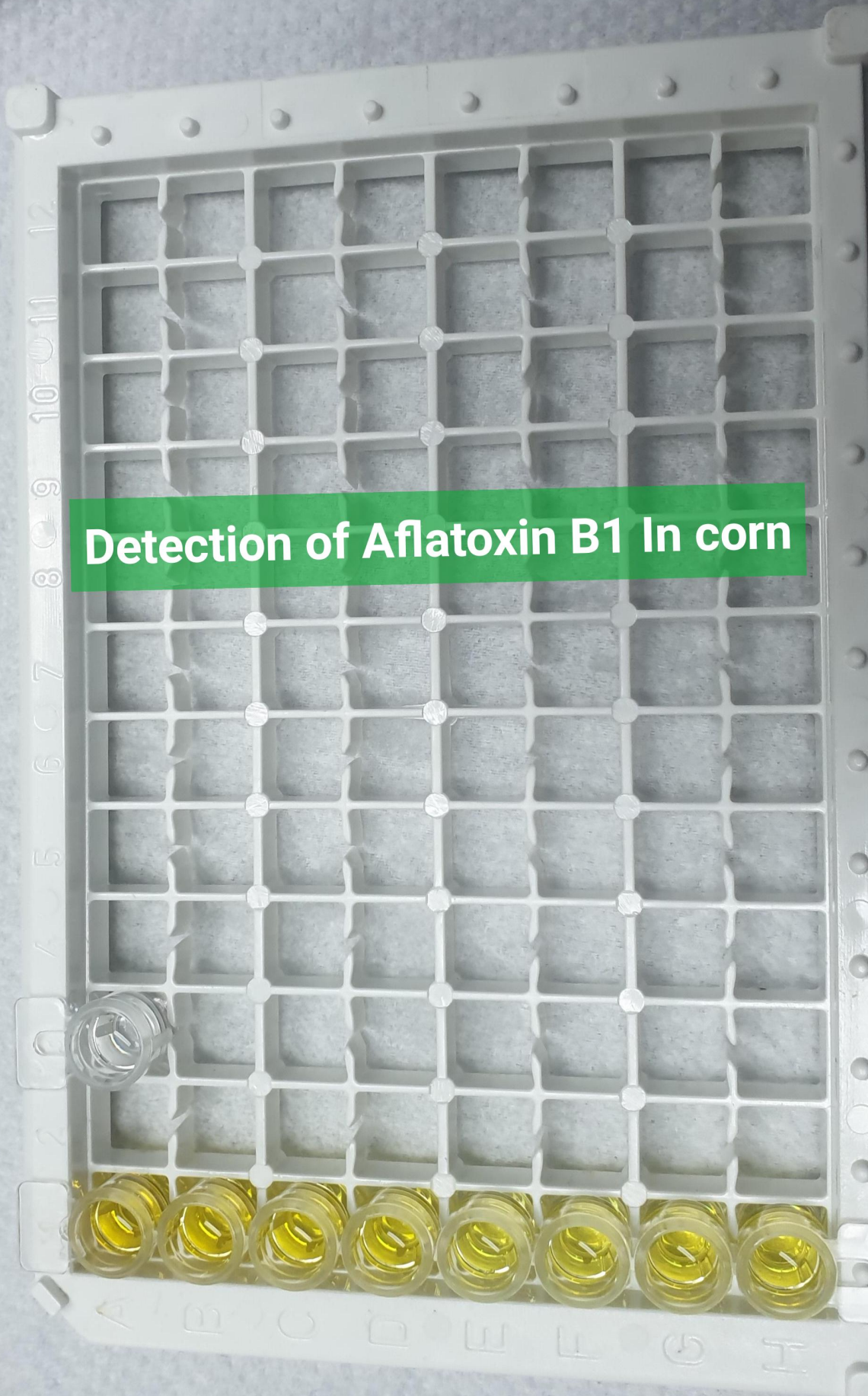
SAMPLE PREPERATION FOR CORN

- weigh 5 g of ground and homogenized sample into a suitable container
- add 25 ml of 70 % methanol.
- shake vigorously for three minutes (manually or with shaker)
- filter the extract through Whatman No. 1 filter (or equivalent) or centrifuge (10 min / 3500 g / room temperature)
- dilute 1 ml of the obtained filtrate or clear supernatant with 1 ml of distilled or deionized water
- use 50 µl of the diluted filtrate per well in the test.

Elisa Test Procedure:-

- Insert a sufficient number of wells into the microwell holder for all standards and samples to be run. Record standard and sample positions.
- Pipet 50 µl of standard or prepared sample into separate wells. Use a new pipette tip for each standard or sample.
- Add 50 µl of conjugate to each well.
- Add 50 µl of antibody to each well.
- Mix gently by shaking the plate manually.
- incubate for 30 min (+/- 1) at room temperature
- Pour the liquid out of the wells and tap the microwell holder vigorously upside down against absorbent paper (three times in a row) to ensure complete removal of liquid from the wells. Fill all the wells with 250 µl wash buffer. Empty the wells again. Repeat two more times.
- Add 100 µl of substrate/chromogen to each well.
- Mix gently by shaking the plate manually and incubate for 15 min (+/- 1) at room temperature.
- Add 100 µl of stop solution to each well.
- Mix gently by shaking the plate manually and measure the absorbance at 450 nm. Read within 15 minutes after addition of stop solution.

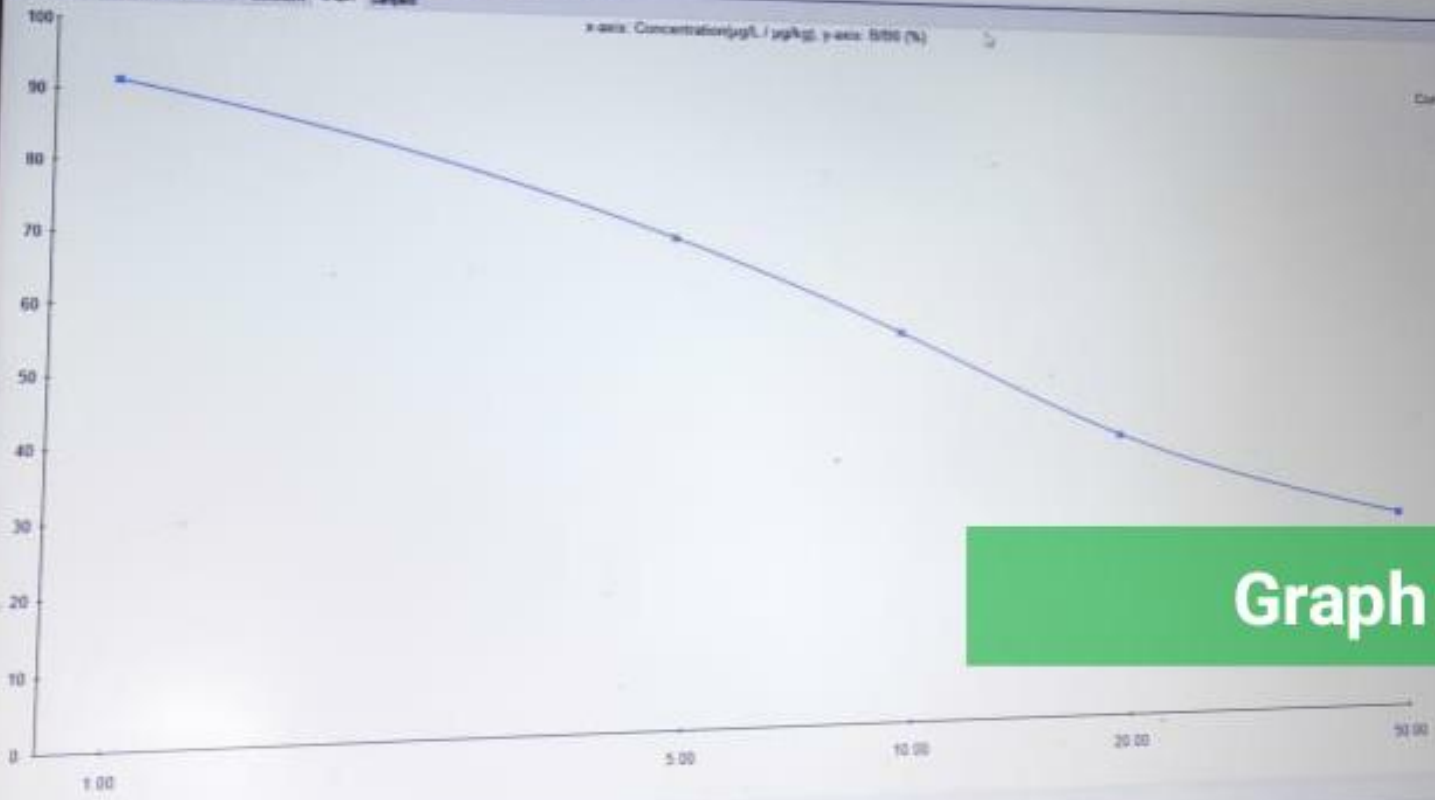
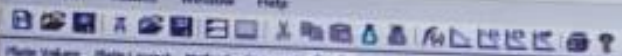
Detection of Aflatoxin B1 In corn



	1	2	3	4	5	6
A	S1 2,614		0003 0.191			
B	S2 2,449					
C	S3 1,986					
D	S4 1,756					
E	S5 1,081					
F	S6 1,252					
G	0001 1,549					
H	0002 1,449					

7-12>> Send Result Print Exit

Reading of the standard and solution



Spine

Conc. (µg/L / µg/kg)	A (Mean)
1.00	2.237 (Coeff. of Var. 0.0%)
5.00	1.789 (Coeff. of Var. 0.0%)
10.00	1.437 (Coeff. of Var. 0.0%)
20.00	1.045 (Coeff. of Var. 0.0%)
50.00	0.737 (Coeff. of Var. 0.0%)
50% addition	13.1

Graph



			Standards			
Ser. No.	Concentration µg/L / µg/kg	Absorbance (Mean) (CV)	B/B0 (%)	calculated µg/L / µg/kg	Deviation (%)	
1	0.00	2.614E 0.0	100.0			
2	1.00	2.449E 0.0	93.7	0.999	0.1	
3	5.00	1.986E 0.0	76.0	5.12	2.4	
4	10.00	1.756E 0.0	67.2	9.84	1.6	
5	20.00	1.081E 0.0	41.4	1129.04	5545.2	
6	50.00	1.252E 0.0	47.9	690.51	1281.0	

			Samples			
Ser. No.	ID	Absorbance (Mean) (CV) (%)	calculated µg/L / µg/kg		µg/L / µg/kg	
1		1.549E 0.0 59.3	12.30	1.00	12.30	
2		1.448E 0.0 55.4	13.51	1.00	13.51	
3		0.191E 0.0 7.3	> 50.00	1.03	> 50.00	

UP-TO-DATE TECHNOLOGIES IN RAPID MICROBIAL TESTING

22nd and 23rd February 2023

Morning
Centenary Block
College

Afternoon
Microbiology Department
Nizam College

The Department
Nizam College



Microbiology

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WELCOME TO 

**TWO DAYS WORKSHOP
ON LATEST TECHNOLOGIES IN
RAPID MICROBIAL TESTING**

22nd and 23rd February 2023

Morning Lectures Centenary Block Nizam College	Afternoon Microbiology Nizam College
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The Program will be held at
Nizam College

