# GOVT. DEGREE COLLEGE FOR WOMEN BEGUMPET, HYDERABAD-16 AUTONOMOUS



# DEPARTMENT OF BIOTECHNOLOGY CBCS SYLLABUS FOR THE YEAR 2021-22



Annexure-1 Credits- for B.Sc. Life Sciences

Courses		Papers	pers Total	Creditsforeachpaper/Semester					
		rapers	Credits	B.Sc.					
Core CoursesDSC	Optional-1	4	30	1	II	111	IV	V	VI
	Optional-2	4	20	5	5	5	5	•	
	Optional-3	4	20	5	5	5	5		•
ElectiveCoursesDSE	Optional-1	1 2	10	5	5	5	5	•	
	Optional-2	2	10	-	-	-	•	5	5
	Optional-3	2	10	•	-	-	-	5	5
Language	English (FirstLanguage)	5	20	4	-	-	-	5	5
Sc.	SecondLanguage	5	20	4	4	3	3	3	3
AbilityEnhancementCom pulsoryCourse	EnvironmentalScience/Basic ComputerSkills	1	20	2	4	3	3	3	3
AECC	BasicComputerSkills/E nvironmentalScience	1	2		2		-		
SkillEnhancementCourse	SEC1	1	2			2		-	
SEC	SEC2	1	2		-	2			
	SEC3	1	2			-	2		
	SEC4	1	2			-	2		
GenericElective GE	OpenStream	1	4	-		-	-	4	•
ProjectWork/Optionals		1	4						4
Total Credits in each semeste	er			25	25	25	25	25	25
TotalCreditsinUG							150	23	
CreditsunderNon-CGPA		NSS/NCC /sports/ Extracurr icular	6	Upto6(2ineachyear)					
		Summer Internship	4	Upto4(2ineach,afterl& II years)					

## Annexurell ProposedNew Grading System

S.No.	GradePoint Rangeofmarks		GradeLetter	
1	10	Equaltoandabove90Marks	A+	
2	9	Morethanorequalto80 andlessthan90Marks	A	
3	8	Morethanorequalto70 andlessthanS0Marks	B+	
4	7	Morethanorequalto60 andlessthan70Marks	В	
5	6	Morethanorequalto55 andlessthan60Marks	C+	
6	5	Morethanorequalto50 andlessthan55Marks	С	
7.	4	Morethanorequalto40 andlessthan50Marks	D	
8	0	Below40 Marks	F	

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# GOVERNMENT DEGREE COLLEGE FOR WOMEN, BEGUMPET, HYDERABAD. AUTONOMOUS (CBCS) PROPOSED SCHEME FOR CHOICE BASED CREDIT SYSTEM FOR B.SC.

## BIOTECHNOLOGY COURSE

9	TECHNOLOGY CO	URSE		
CODE	COUDER TO	ECTED A		
	COURSETITIE			
BS 101	Environmental	COURSE	H PW	CREDITS
BS 102	Environmental Science/Basic Computer Skills English	TYPE		
BS 103	Second I-	AECC-1	2	
BS 104	1	CC-1A	4	4
BS 105	Trional 1- ( PH biological)	CC-2A	4	4
BS 106		DSC-1A	4T+3P=7	4+1=5
-0100	Optional III	DSC-2A		4+1=5
	TOTAL	DSC-3A		4+1=5
DC 201	TIVE CONTRACTOR OF THE PROPERTY OF THE PROPERT	COMPANY AND		25
BS 201	Gender Sensitization   FIRST YEAR-SEME			
BS 202	English State	AECC-2	2	2
BS 203	Second language	CC-1B	4	4
BS 204	Optional II Optional II	CC-2B	4	4
BS 205	Optional II	DSC-1B	4T+3P=7	4+1=5
BS 206	Optional III	DSC-2B		4+1=5
		DSC-3B		4+1=5
	TOTAL		Z.,	25
7.4	SECOND YEAR- SEME	STER III		
BS 301	SEC 1: UGC specified SEC	SEC-1	47 5 (2.1	
BS 302	SEC 2: Immunological techniques	SEC-1	2	Lind No. THE
BS 303	English	CC-1C	2	2
BS 304	Second language	CC-2C	3	3
BS 305	Optional I- Molecular Biology and			3
DO 202	Recombinant DNA Technology	DSC-1C	4T+3P=7	4+1=5
BS 306	Optional II	DSC-2C		4+1=5
BS 307	Optional III	DSC-3C		4+1=5
וטע טע	TOTAL	D50-50		-
	SECOND YEAR- SEME	STED IV	1	25
1, -		· 1		
BS 401 BS 402	SEC 3: UGC specified SEC	SEC-3	-	1
BS 402	SEC 4: Molecular markers in plant breeding	SEC-4 CC- I D	3	3
3S 403	English			
S 404	Second language	CC-2D	3	3
3S 405	Optional I- Bioinformatics and Biostatistics	DSC-1D	4T+3P=7	4+1=5
	Optional II	DSC-2D		4+1=5
S 406	Optional III	DSC-3D		4+1=5
3S 407	TOTAL			25
				*
	Markey Community And	1.		

## PROPOSED SCHEME FOR CHOICE BASED CREDIT SYSTEM FOR B.SC. BIOTECHNOLOGY COURSE

FIRST YEAR- SEM	ESTER I		
COURSE TITLE		H PW	CDEDITO
Environmental Science (P. 1)	TYPE	1111	CREDITS
English English	AECC-1	2	
	CC-1A		4
Ontional L. Call L.	CC-2A	4	4
Optional II	DSC-1A	4T+3P=7	4+1=5
Optional III	DSC-2A		4+1=5
	DSC-3A		4+1=5
			25
Gandon San iti si	STER II		
	AECC-2	2	2
	CC-1B	4	4
Second language	CC-2B	4	4
Optional I- Biological Chemistry and Microbiology	DSC-1B	4T+3P=7	4+1=5
Optional II	DSC-2B		4+1=5
1	DSC-3B	, ,	4+1=5
TOTAL			25
SECOND YEAR-SEME	STER III		
		1 2 2 2	
		2	2
			3
			3
			4+1=5
	DSC-1C	41131-7	411-3
	DSC-2C		4+1=5
	DSC-3C		4+1=5
			25
	STER IV		
SEC 3: UGC specified SEC	SEC-3		
SEC 4: Molecular markers in plant breeding	SEC-4	2	2
	CC-1D	3	3
Casand language	CC-2D	3	3
Second language	DSC-1D	4T+3P=7	4+1=5
Ontional L. Bioinformatics and Biostatistics	DSC-1D		
Optional I- Bioinformatics and Biostatistics		12.02	4+1=5
Optional I- Bioinformatics and Biostatistics Optional II	DSC-2D		4+1=5 4+1=5
Optional I- Bioinformatics and Biostatistics			
	Environmental Science/Basic Computer Skills English Second language Optional II- Cell biology and Genetics Optional III  TOTAL  FIRST YEAR-SEME  Gender Sensitization   Computer Skills English Second language Optional II- Biological Chemistry and Microbiology Optional III  TOTAL  SECOND YEAR-SEME  SEC 1: UGC specified SEC  SEC 2: Immunological techniques English Second language Optional I- Molecular Biology and Recombinant DNA Technology Optional III  Optional III  TOTAL  SECOND YEAR-SEME  SEC 3: UGC specified SEC  SEC 4: Molecular markers in plant breeding English	Environmental Science/Basic Computer Skills  English  Second language  Optional I- Cell biology and Genetics  Optional III  DSC-2A  Optional III  FIRST YEAR-SEMESTER II  Gender Sensitization  FIRST YEAR-SEMESTER II  Gender Sensitization  CC-1B  Second language  Optional I- Biological Chemistry and Microbiology  Optional III  DSC-2B  Optional III  DSC-2B  Optional III  DSC-2B  Optional III  SEC 1: UGC specified SEC  English  SEC-1  SEC 2: Immunological techniques  SEC-2  English  CC-1C  Second language  Optional I- Molecular Biology and  Recombinant DNA Technology  Optional III  DSC-2C  Optional III  DSC-2C  Optional III  DSC-3C  SEC-3  SEC 3: UGC specified SEC  SEC-3  SEC 4: Molecular markers in plant breeding  SEC-4  English  CC-1D	COURSE TITLE  Environmental Science/Basic Computer Skills  English  Second language  Optional I- Cell biology and Genetics Optional II  Optional III  DSC-2A  Optional III  FIRST YEAR-SEMESTER II  Gender Sensitization  FIRST YEAR-SEMESTER II  Gender Sensitization  Computer Stelly  FIRST YEAR-SEMESTER II  Gender Sensitization  Computer Stelly  FOR Second language  CC-2B  Optional II DSC-3A  Optional II DSC-3B  Optional II DSC-3B  TOTAL  SECOND YEAR-SEMESTER III  SEC 1: UGC specified SEC  English  CC-1C  Second language  CC-2C  3  Optional I- Molecular Biology and PSC-1C  Recombinant DNA Technology  Optional III  DSC-3C  Optional III  DSC-3C  SEC-3  SEC-4  AT+3P=7  Recombinant DNA Technology  Optional III  DSC-3C  SEC-3  SEC-4  SEC-6  SEC-6  SEC-6  SEC-7  SEC-7  SEC-8  SEC-9  SE

CODE	COURSE TITLE	COURSE	HPW	CREDITS
BS 501	English	CC-1E	ò	3
BS 502	Second language	CC-2E	3	3
BS 503	Basics in Biotechnology	GE	4	4
BS 504	Optional I- A/B (A) Plant Biotechnology or (B) Medical Biotechnology	DSE -1E	4T+3P=7	4+1=5
BS 505	Optional- II A/B	DSE -2E		4+1=5
BS 506	O.C. L. TITLA ID	DCC 2C	15 15	4+1=5
20 200	Optional- III A/B	DSE -3E		1 411-5
28 300	TOTAL			25
	TOTAL  THIRD YEAR- SEMESTER  Project in Biotechnology/	R VI Project	4	
BS 601	TOTAL  THIRD YEAR- SEMESTER  Project in Biotechnology/ Optional I: (IPR, Biosafety and Entrepreneurship)	R VI	4 3	25
BS 601	TOTAL  THIRD YEAR- SEMESTER  Project in Biotechnology/ Optional I: (IPR, Biosafety and Entrepreneurship)  English	Project work/Opt.P		25
BS 601 BS 602 BS 603	TOTAL  THIRD YEAR- SEMESTER  Project in Biotechnology/ Optional I: (IPR, Biosafety and Entrepreneurship)	Project work/Opt.P CC-1F	3	4 3
BS 601 BS 602 BS 603 BS 604	TOTAL  THIRD YEAR- SEMESTER  Project in Biotechnology/ Optional I: (IPR, Biosafety and Entrepreneurship)  English  Second language  Optional II- A/B  (A) Animal Biotechnology or (B) Environmental Biotechnology	Project work/Opt.P CC-1F CC-2F	3 3	4 3 3
BS 601 BS 602 BS 603 BS 604	TOTAL  THIRD YEAR- SEMESTER  Project in Biotechnology/ Optional I: (IPR, Biosafety and Entrepreneurship)  English  Second language  Optional II- A/B  (A) Animal Biotechnology or (B) Environmental Biotechnology  Optional- II A/B	Project work/Opt.P CC-1F CC-2F DSE-1F	3 3	25 4 3 3 4+1=5
BS 601 BS 602 BS 603 BS 604 BS 605 BS 606	TOTAL  THIRD YEAR- SEMESTER  Project in Biotechnology/ Optional I: (IPR, Biosafety and Entrepreneurship)  English  Second language  Optional II- A/B  (A) Animal Biotechnology or (B) Environmental Biotechnology	Project work/Opt.P CC-1F CC-2F DSE-1F	3 3	25 4 3 3 4+1=5 4+1=5

Total credits= 164-12 (AECC 4 + SEC 8) =15

AECC: Ability Enhancement Compulsory Course

SEC: Skill Enhancement Course

SEC\*:SEC(UGC Recommended Courses)

DSC: Discipline Specific Course DSE: Discipline Specific Elective

GE: Generic Elective

been changed to two UGC recommended SEC courses. SEC 1: Industrial Fermentation\* and SEC 4: Drug designing\*

## GOVERNMENT COLLEGE FOR WOMEN BEGUMPET, HYDERABAD - 500016 (AUTONOMOUS) Constitution of Board of Studies

S.No	Name of Board	of Studies - 2018-2021	
01.	V.Rohini,		Signature
	Assistant professor of Biotechnology.	Chairman BOS, GDC(w) Begumpet	Police
02.	Dr. Smita C. Pawar Head, Department of Genetics, Osmania university	Chairperson BOS, University Nominee, Osmania University	ABSENT
03.	Dr. A.Uma Associate Professor, BOS chair- Centre for Biotechnology, Officer - Incharge Examination, Institute of Science and Technology, JNTUH	Subject Expert Nominated by Academic Council	fre
04.	Dr.Anil Pasupulati Assistant Professor, Department of Biochemistry, School of Life Sciences, University of Hyderabad	Subject Expert Nominated by Academic Council	p. Attau
05	Mr.Mahesh Kyasani, Sr Manager, Manufacturing Sciences, Biological E Limited, Shameerpet	Representative from Industry	Mahuh
)6 	Dr.M.Vasudha, Assistant Professor of Genetics, Government Degree College for Women, Begumpet, Hyderabad.	Member	Vasadha M
7	<b>D.Soumya</b> , M.Sc. Biotechnology, University college Science, Saifabad Hyderabad	Alumnus	Depunya

## Government Degree College for Women, Begumpet. Hyderabad. Autonomous(CBCS)

Agenda for the Board of Studies Meeting (2021-22)

- 1. Approval of the Syllabus for V and VI Semesters of the B.Sc., III year according to CBCS.
- 2. Review of the Syllabus and Model papers.
- 3. Preparation of Panel of Examiners and their approval
- 4. Review and suggestions on Methodologies for innovative teaching and evaluation techniques and extension activities.
- 5. Reapproval of the syllabus for I and II semesters of B.Sc., I Year according to CBCS.
- 6. Approval of scheme of evaluation (60 marks for the External examination and 40 marks for the Internal examination)

Division of 40 marks is as follows.

- 1. 20marks internal assessment in the form of descriptive exam, where two internals will be conducted and average of two is considered.
- 2. Unit wise test in the form of 20 objective questions, half mark each and a total value of 10 marks.
- 3. 5marks for Seminar/Quiz / group discussion and 5 marks for assignments.

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## Government Degree College for Women, Begumpet. Hyderabad. Autonomous(CBCS)

## Board of Studies Meeting (2021-22)

Keeping in view of the CBCS Pattern, rearrangement of B.Sc. III yr syllabus was discussed at length for the approval with the members of BOS Committee.

The ratification of the existing pattern of marks distribution of B.Sc. III yr Semester-V&VI Biotechnology syllabus has been approved by the members in the BOS Committee.

Marks distribution for Semester-V & VI is

Theory-Internal Assessment- 40 Marks

- > (2 Internals-20+20, Average of the two internals will be taken)+5M Assignment (write up of 4-5 pages)+5M Seminar+10 M MCQS(Average of 4 MCQS Tests)
- External Examination -60 Marks

Total: 100 Marks

- Practical External Examination: 50 Marks
- > SEMESTER -V, Title of the Paper-V (DSE-I) is" PLANT BIOTECHNOLOGY" (Theory& Practical)
- > Paper V (GE-I) BASICS OF BIOTECHNOLOGY- (Theory)
- > SEMESTER -VI, Title of the Paper-VI (DSE-II) is "ANIMAL BIOTECHNOLOGY" (Theory& Practical)
- > PROJECT- 100 Marks (-4 credits)

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# GOVERNMENT DEGREE COLLEGE FOR WOMEN (UG & PG) AUTONOMOUS (CBCS) BEGUMPET, HYDERABAD-500016 DEPARTMENT OFBIOTECHNOLOGY.

The Panel of Examiners nominated and approved by the Board of Studies members for the Semesters Examination of Biotechnology has been enlisted below.

S.No	Name	Designation	Address
1	Dr. H.Surekha Rani, Mobile: 9866620067	Assistant Professor of Biotechnology	Department of Genetics and Biotechnology Osmania University
2	Dr. A.Uma Mobile: 9848120819 Vedavathi1@jntuh.ac.in	Associate Professor of Biotechnology BOS chair-	Centre for Biotechnology, IST - JNTUH
3.	Dr.Anil Pasupulati anilkumar@uohyd.ac.in	Assistant Professor, Department of Biochemistry	School of Life Sciences, University of Hyderabad
4	Dr. Y. Venkateswarlu Mobile: 8801150220	Asst. Professor of Biotechnology	Govt. Degree college, Khairatabad ,Hyderabad
5	Dr. P. Suresh Kumar Mobile: 9701407475	Head ,Asst. Professor of Biotechnology	Loyola Academy Degree and P.G College, Hyderabad
6	Ms. D.Annapurna Mobile 9959220195	Asst. Professor of Biotechnology	Tara GDC,Sanga Reddy
7.	Mrs. P. Pushpalatha Mobile: 8099872878	Head ,Asst. Professor of Biotechnology	Govt. City college, Hyderabad.
8	Ms.D.Kethora	Asst. Professor of Biotechnology	St.Ann's Degree College for Women, Mehdipatnam, Hyd
9	Dr.D.Sambhasiva	Assistant Professor of Biotechnology	NizamCollege,Hyderabad

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#### B. Sc. I Year (2020-21) MODEL QUESTION PAPER

Subject: Biotechnology

Time: 2 hrs

Max Marks: 60 Min marks: 24

SECTION-A

I Write any 5 Short Answer Type Question 5×4=20M

(2 from each unit)

- 1.
- 2.
- 3,
- 4.
- 5.
- 6. 7.
- 8.

#### SECTION-B

II Essay Type Questions (Internal Choice)

4×10≈40M

- I. UNIT I
- (or) В
- 2. UNIT II :
- A (or) B
- 3. UNIT III :
- (or) B
- 4. UNIT IV :
- (or) В

B. Sc. (2021-22)

## INTERNAL ASSESSMENT MODEL QUESTION PAPER

Subject: Biotechnology Max.MARKS:20

2x5=10M
1x10=10M

## PROGRAM OUTCOMES

## PO 1 Domain Expertise:

- Acquire comprehensive knowledge and skills.
- Make use of the knowledge in an innovative manner.
- Effectively apply the knowledge and skills to address various issues.

## PO 2 Modern equipment Usage

- Use ICT effectively.
- Access, retrieve and use authenticated information.
- Access, retrieve and use authenticated information. Have knowledge of software applications to analyze data.

## PO 3 Computing Skills and Ethics

- Develop rationale and scientific thinking process.
- Use technology intelligently for communication, entertainment and for the benefit of mankind.
- Ensure ethical practices throughout ones endeavors for the wellbeing of human race.

## PO 4 Complex problem Investigation & Solving

- Predict and analyze problems.
- Frame hypotheses.
- Investigate and interpret empirical data.
- Plan and execute action.

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## PO 5 Perform effectively as Individuals and in Teams

- Work efficiently as an individual
- Cooperate, coordinate and perform effectively in diverse teams/groups.
- Prioritize common interest to individual interest.

## PO 6 Efficient Communication & Life Skills

- Express thoughts in an effective manner
- Listen, understand and project views in a convincing manner.
- Decide appropriate media to share information
- Develop skills to present significant information clearly and concisely to interested groups.

## PO 7 Environmental Sustainability

- Understand sensibly the Environmental challenges.
- Think critically on environment sustainability measures.
- Propagate and follow environment friendly practices.

#### PO 8 Societal contribution

- Render service for the general good of the society.
- Involve voluntarily in social development activities at Regional, National, global levels.
- Have own pride in volunteering to address societal issues viz: calamities, disasters, poverty, epidemics.
- Be a patriotic citizen to uphold the values of the nation

### PO 9 Effective Project Management

- Identify the goals, objectives and components of a project and decide the appropriate time of completion.
- Plan, organize and direct the endeavors of teams to achieve the set targets in time.
- Be competent in identifying opportunities and develop strategies for contingencies.

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## Programme Specific Outcomes of B. Sc. in Biotechnology

- 1. Apply the knowledge of various branches of Biotechnology to solve the complex problems.
- 2. Identify, formulate, research literature, and analyze problems to arrive at substantiated conclusions natural sciences.
- 3. Design solutions for the public health and safety, and the cultural, societal, and environmental considerations.
- 4. Use research-based knowledge including design of experiments, analysis and interpretation of data, and synthesis of the information to provide valid conclusions.

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B.Sc. Biotechnology I YEAR (2021-22) SEMESTER- 1 DSC-Paper- I: CELL BIOLOGY AND GENETICS

#### COURSE OUTCOMES

#### CREDITS-4 TEACHING HOUR/WEEK-4

After completion of the course student will be able to:

- Recall the history of cytology and distinguish between Prokaryotic and Eukaryotic cell
- Explain the organization of Genes and chromosomes, chromosome morphology and its aberrations
- Compare and contrast the events of cell cycle and its regulation.
- The student will demonstrate proficiency in understanding the basic structure of cell and interpret the inheritance of characters by using linkage and crossing over.

#### **Unit 1: Cell structure and Functions**

- Seminar

1.1. Cell as basic unit of living organisms-bacterial, fungal, plant and animal cells

1.2. Ultrastructure of prokaryotic cell (cell membrane and plasmids, Nucleoid)

1.3. Ultrastructure of eukaryotic cell (cell wall, cell membrane, nucleus, mitochondria, chloroplast, endoplasmic reticulum, Golgi apparatus, vacuoles)

1.4. Fluid mosaic model, Sandwich model, Cell membrane permeability

1.5. Structure of chromosome-morphology, components of chromosomes (histones and nonhistones),

1.6. Specialized chromosomes (Polytene, Lampbrush) Structural and Numerical Aberrations

#### Unit 2: Cell cycle

- 2.1 Bacterial cell division
- 2.2 Eukaryotic cell cycle -phases
- 2.3 Mitosis Stages -significance
- 2.4 Meiosis- Stages-significance
- 2.5 Senescence and necrosis
- 2.6 Apoptosis

## Unit 3: Principles and mechanism of inheritance

3.1. Mendel's experiments - factors contributing to success of Mendel's experiments

3.2. Law of segregation - Monohybrid Ratio; Law of independent assortment- Dihybrid ratio, Trihybrid ratio

3.3. Deviation from Mendel's laws- partial or incomplete dominance (eg: Flower Color in Mirabilis jalapa), Co-dominance (eg: MN Blood groups), Non allelic interactions - types of epistasis, modification of dihybrid ratios(12:3:1; 9:7; 15:1; 9:3:4,9:7; 13:3)

3.4. Penetrance and Expressivity (eg: Polydactyly. Waardenburg syndrome).pleiotropism, phenocopymicrocephaly, cleft lip.

3.5. Multiple alleles (eg: Coat color in Rabbits, eye color in Drosophila and ABO Blood groups)

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3.6. X-Y chromosomes - Sex determination in Drosophila, Man, X-linked inheritance - Hemophilia and Color blindness; X-inactivation.

## Unit 4: Linkage, Recombination and Extension to Mendel's Laws

4.1. Linkage and recombination - Cytological proof of crossing over, phases of linkage, recombination frequency, gene mapping and map distance

4.2. Non-Mendelian Inheritance - Maternal effect (Shell coiling in snail), variegation in leaves of Mirabilis jalapa

4.3. Cytoplasmic male sterility in Maize.

- 4.4. Mitochondrial inheritance in human and poky in Neurosporacrassa
- 4.5. Chloroplast inheritance in Chlamydomonas

4.6. Hardy-Weinberg Equilibrium.

#### OPTIONAL- I: PRACTICALS CELL BIOLOGY AND GENETICS

- 1. Microscopic observation of cells: bacteria, fungi, plant and animal
- 2. Preparation of different stages of Mitosis (onion root tips)
- 3. Preparation of different stages of Meiosis (grasshopper testis)
- 4. Preparation of Polytene chromosome from Drosophila salivary gland
- 5. Monohybrid and dihybrid ratio in Drosophila
- 6. Monohybrid and dihybrid ratio in Maize
- 7. Problems on co-dominance, epistasis, two point and three point test cross, gene mapping.
- 8. Statistical applications of Hardy-Weinberg Equilibrium

#### Spotters:

- 1. Prokaryotic Cell (Bacteria)
- 2. Mitochondria
- 3. Chlorolplast
- 4. Polytene Chromosomes
- 5. Test Cross
- 6. Blood Grouping
- 7. Hemophilia Pedigree
- 8. Crossing Over
- 9. Synaptonemal Complex
- 10. Nucleosome Model

#### REFERENCE BOOKS

- 1. Cell & Molecular Biology. E.D.D De Robertis&E.M.F De Robertis, Waverly publication
- 2. An introduction to Genetic Analysis by Anthony, J.F. J.A. Miller, D.T. Suzuki, R.C. Richard Lewontin, W.M-Gilbert, W.H. Freeman publication
- 3. Principles of Genetics by E.J.Gardner and D.P. Snusted. John Wiley & Sons, New York
- 4. The science of Genetics, by A.G. Atherly J.R. Girton, J.F. Mcdonald, SaundernCollege publication

5. Principles of Genetics by R.H. TamarinMcGrawhill

6. Theory & problems in Genetics by Stansfield, Schaum out line series McGrawhill

7. Molecular Cell Biology Lodish, H., Baltimore, D; fesk, A., Zipursky S.L., Matsudaride, P. and Darnel. American Scientific Books. W.H. Freeman, New York

8. The cell: A molecular approach. Geoffrey M Cooper, Robert E Hausman, ASM press

9. Cell and Molecular Biology, Concepts and Experiments — Gerald Karp, John Wiley &Sons, Inc.

10.Cell Biology And Genetics by P.K. GUPTA

BSC BIOTECHNOLOGY-I YEAR (2021-22)

SEMESTER- II DSC-Paper- II: BIOLOGICAL CHEMISTRY AND MICROBIOLOGY

#### COURSE OUTCOMES

**CREDITS-4 TEACHING** 

#### HOUR/WEEK-4

After completion of the course student will be get exposed

- To strong theoretical and practical background in fundamental concepts.
- To get insights of multiple important technical areas of Biochemistry.
- To apply contextual knowledge and modern tools of biochemical research for solving problems.
- To give students a generalized idea about microbiology its basic aspects

### **Unit 1: Biomolecules**

- 1.1 Carbohydrates- importance, classification; structure and functions of monosaccharaides (glucose& fructose), disaccharides (sucrose, lactose & maltose) and polysaccharides (starch, glycogen & inulin)
- 1.2 Amino acids- importance, classification, structure, physical and chemical properties of amino acids; peptide bond formation
- 1.3 Proteins- importance, structure of proteins- primary, secondary, tertiary and quaternary
- 1.4 Lipids- importance, classification- simple lipids (triacylglycerides& waxes), complex lipids (phospholipids & glycolipids), derived lipids (steroids, terpenes& carotenoids)
- 1.5 Nucleic acids :structure and chemistry of DNA (Watson and crick) and RNA(TMV) Structure and forms of DNA (A, B and Z)
- 1.6 Enzymes- importance, classification and nomenclature; Michaelis-Menton Equation, factors influencing the enzyme reactions; enzyme inhibition (competitive, uncompetitive & mixed), co-enzymes

## **Unit 2: Bioenergetics**

- 2.1 Glycolysis, Tricarboxylic Acid (TCA) Cycle,
- 2.2 Electron Transport, Oxidative Phosphorylation
- 2.3 Gluconeogenesis and its significance
- 2.4 Transamination and Oxidative deamination reactions of amino acids
- 2.5 B-Oxidation of Fatty acids
- 2.6 Glyoxalate cycle

## 3. Unit: Fundamentals of Microbiology

3.1 Historical development of microbiology and contributors of microbiology

- 3.2 Microscopy: Bright field microscopy, Dark field microscopy, Phase contrast microscopy, Fluorescent microscopy, Scanning and Transmission electron microscopy
- 3.3 Outlines of classification of microorganisms
- 3.4 Structure and general characteristics of bacteria and virus
- 3.5 Disease causing pathogens and symptoms (Eg: Mycobacterium, Hepatitis)
- 3.6 Structure and general characteristics of micro-algae and fungi

## 4. Unit: Culture and identification of microorganisms

- 4.1 Methods of sterilization- physical and chemical methods
- 4.2 Bacterial nutrition nutritional types of bacteria, essential macro& micro nutrients and growth
- 4.3 Bacterial growth curve-batch and continuous cultures, synchronous cultures measurement of bacterial growth-measurement of cell number and cell mass.
- 4.4 Factors affecting bacterial growth
- 4.5 Culturing of anaerobic bacteria and viruses
- 4.6 Pure cultures and its characteristics

#### OPTIONAL I: PRACTICALS

#### BIOLOGICAL CHEMISTRY AND MICROBIOLOGY

- 1. Preparation of normal, molar&molal solutions.
- 2. Preparation of buffers (acidic, basic& neutral)
- 3. Qualitative tests of sugars, amino acids& lipids
- 4. Estimation of total sugars by anthrone method
- 5. Separation of amino acids by paper chromatography
- 6. Estimation of proteins by biuret method
- 7. Sterilization methods
- 8. Preparation of microbiological media (bacterial, algal & fungal)
- 9. Isolation of bacteria by streak, spread and pour plate methods
- 10. Isolation of bacteria from soil
- 11 Simple staining and differential staining (gram's staining)
- 12 Bacterial growth curve
- 13 Technique of micrometry (ocular and stage)

#### Spotters:

- 1. Osazone
- 2. Globular protein
- 3. Lock and key, model
- 4. Complete inhibition
- 5. RUBISCO
- 6. ATP synthase
- 7. Autoclave
- 8. Laminar air flow
- 9. Tyndalization



- 10. Bacterial growth curve
- 11. Hot air oven
- 12. Serial dilution technique

### REFERENCE BOOKS

- 1. Lehninger Principles of Biochemistry By: David L. Nelson and Cox
- 2. Biochemistry By: Rex Montgomery
- 3. Harper's Biochemistry By: Robert K. Murray
- 4. Enzymes By: Trevor Palmer
- 5. Enzyme structure and mechanism By: AlanFersht
- 6. Principles of Biochemistry By: Donald J. Voet, Judith G.Voet, Charlotte W.Pratt
- 7. Analytical Biochemistry By: Cooper
- 8. Principles and techniques of Biochemistry and Molecular Biology Edited By: Keith Wilson
- 9. Practical Biochemistry By: Plummer
- 10. Biology of Microorganisms by: Brock, T.D. and Madigan, M.T.
- 11. Microbiology by: Prescott, L.M., Harley, J.P. Klein, D.A.
- 12. Microbiology by: Pelczar, M.J, Chan, E.C.S., Ereig, N.R.
- 13. Microbiological applications by: Benson

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## BSC BIOTECHNOLOGY-II YEAR (2021-22)

Semester III- Molecular Biology and Recombinant DNA Technology

#### COURSE OUTCOMES

#### CREDITS-4 TEACHING HOUR/WEEK-

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After completion of the course student will understand:

- Structural levels of nucleic acids- DNA and RNA and genome organization in prokaryotes and eukaryotes.
- The concept of Gene and the gene architecture.
- Molecular Events of Transcription and processing of transcripts
- The knowledge of recombinant DNA technology

#### Unit 1: Nucleic Acids and Genome organization

- 1.1 DNA as the genetic material- Griffith's experiments on transformation, Avery McCleod and McCarty experiment, Hershey-Chase experiment, RNA as Genetic Material
  - 1.2 Genome organization in prokaryotes and Eukaryotes
  - 1.3 Genome organization in Mitochondria and Chloroplast genome
  - 1.4 DNA replication- Semi conservative DNA replication-Messelson and Stahl experiment
  - 1.5 Replication in Prokaryotic Genome and Nuclear Genome of Eukaryotes
  - 1.6 Mutation-Spontaneous and Induced, Physical and chemical Mutagens

#### 2. Gene expression in prokaryotes and Eukaryotes

- 2.1 Structure of prokaryotic and Eukaryotic gene ,Structure and functions of prokaryotic RNA polymerase
- 2.2 Transcriptional machinery of eukaryotes-Structure and functions of eukaryotic RNA polymerase
- 2.3 Genetic Code-Properties, deciphering genetic code, wobble hypothesis
- 2.4 Prokaryotic Transcription- initiation, elongation, proof reading and termination (rho dependent and independent),
- 2.5 Eukaryotic Transcription- initiation, elongation and termination
- 2.6 Prokaryotic and eukaryotic- Translation- initiation, elongation and termination.

#### 3. Unit: Gene regulation in Prokaryotes and Eukaryotes

- 3.1 Prokaryotic transcriptional regulation (inducible System)-Operon concept, Lac operon, glucose effect.
  - 3.2 Prokaryotic transcriptional regulation (repressible system)- Tryptophan operon
  - 3.3 Post transcriptional modifications Capping and Poly adenylation
  - 3.4 Splicing and alternate splicing

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3.5 Post translational modification- glycosylation and adenylation and ubiquitination 3.6 Gal regulation in yeast-mating type gene switching

## Unit 4: Recombinant DNA Technology \_ Sombass.

4.1 Enzymes useful in molecular cloning: Restrictionendonuclease, DNA ligases, Polynucleotide kinase, DNA Polymerase, klenow enzyme, reverse transcriptase, Alkaline phosphatase, terminal nucleotidyltransferase

4.2 Cloning Vectors: pBR322, Bacteriophage, Cosmid, Phagemid, Shuttle vectors

4.3 Vectors for library preparation (lambda phage vector, Cosmid, BAC and YAC)

4.4 Gene transfer techniques: Physical, Chemical and Biological methods

4.5Se; lection of recombinant clones-colony hybridization and library screening

4.6 Polymerase Chain Reaction and Applications of recombinant DNA technologies-Agriculture, Medicine

#### **PRACTICALS** MOLECULAR BIOLOGY AND RECOMBINANT DNA TECHNOLOGY

- Isolation of DNA from bacterial cell
- 2. Isolation of DNA from Plasmid
- 3 Agarose gel electrophoresis of DNA
- 4. Quantification of DNA by Spectrophotometer
- 5 Separation of proteins by SDS-PAGE
  6 Polymerase Chain Reaction
  7 Restriction digestion of DNA

- 8) Bacterial Transformation (Selection of transformants with blue white selection)

#### REFERENCE BOOKS

- 1. Molecular Biology of the cell. Alberts, B; Bray, D, Lews, J., Raff, M., Roberts, K and Watson, J.D. Garland publishers, Oxford
- 2. Molecular Biology of the Gene By Watson, Hopkins, Goberts, Steitz and Weiner (Pearson Education)
- 3. Text Book of Biotechnology By H.K. Das (Wiley Publications)
- 4. Gene Structure & Expression By J.D. Howkins, Publ: Cambridge
- 5. Test Book of Molecular Biology By K.S. Sastry, G. Padmanabhan& C. Subramanyan, Publ: Macmillan India
- 6. Principles of Gene Manipulation By R.W. Old & S.B. Primrose, Publ: Blackwell
- 7. Genes By B. Lewin Oxford Univ. Press
- 8. Molecular Biology &Biotechnol. By H.D. Kumar, Publ: Vikas
- 9. Methods for General & Molecular Bacteriology By P. Gerhardf et al., Publ: ASM
- 10. Molecular Biotechnology By G.R. Click and J.J. Pasternak, Publ: Panima
- 11. Genes and Genomes By Maxine Singer and Paul Berg
- 12. Molecular Biology By D. Freifelder, Publ: Narosa
- 13. Molecular biology. By;F. Weaver. WCB/McGraw Hill.

# GOVERNMENT DEGREE COLLEGE FOR WOMEN, BEGUMPET, HYDERABAD. AUTONOMOUS (CBCS) BSC BIOTECHNOLOGY-II YEAR (2021-22) SEMESTER-III

SKILL ENHANCEMENT COURSE -2 (SEC- 2) BS 302: IMMUNOLOGICAL TECHNIQUES

- 1. Unit: Antibody assays Principle, Methodology and Applications
- 1.1. Antigen Antibody reactions: opsonisation, neutralization, precipitation & agglutination
- 1.2. Immuno diffusion & radial diffusion
- 1.3. Immuno electrophoresis rocket and counter current
- 1.4. ELISA & western blotting
- 1.5. Radioimmunology assay & immune fluorescent assay
- 1.6. Immlmohisto chemistry
- 2. Unit: Cellular Assays Principle, Methodology and Applications
- 2.1. Total and differential count in human peripheral blood
- 2.2. Separation of mononuclear cells from human peripheral blood
- 2.3. Cell viability assay using tryphan blue
  - 2.4. Lymphocyte transformation assay
- 2.5. Enumeration of T & B cells from human peripheral blood
- 2.6. Micro cytotoxicity assay for HLA typing

#### REFERENCE BOOKS

- 1. Essential Immunology by I. Roitt, Publ: Blackwell
- 2. Immunology by G. Reever& I. Todd, Publ; Blackwell
- 3. Cellular and Molecular Immunology by Abbas AK, Lichtman AH, Pillai S. Saunders publication, Philadelphia
- 4. Kuby's Immunology by Golds RA, Kindt TJ, Osborne BA. W.H. Freeman and company, New York

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# GOVERNMENT DEGREE COLLEGE FOR WOMEN, BEGUMPET, HYDERABAD. AUTONOMOUS (CBCS) BSC BIOTECHNOLOGY-II YEAR (2021-22)

Semester IV (DSC – 1D)
B S 405: Bioinformatics and Biostatistics

## COURSE OUTCOMESCREDITS-4 TEACHING HOUR/WEEK-4

After completion of the course student will understand how to:

- Store and Retrieve drug related information using online tools
- Comprehend the utility of tools & databases available in genomic & proteomics
- Understand simple calculations, to plan and execute research designs
- Analyse data, interpret, information and compare observed data

## Unit 1: Introduction to Bioinformatics and Biological Databases

1.1 Bioinformatics – a history, Scope and applications

1.2 Bioinformatics tools and resources, internet basics, role of internet, free online tools, downloadable tools

1.3 Bioinformatics web portals-NCBI, EBI, ExPASy

1.4 Biological databases: classification of Databases primary (Genbank), Secondary (PIR), Tertiary and composite (KEGG) databases

1.5 Sequence Databases – DNA sequence databases

1.6 Protein data sequence databases-(swissprot and PROSITE)

#### Unit 2: Sequence Alignment

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- 2.1 Basics of sequence alignment match, mismatch, gaps, gap penalties, scoring alignment
- 2.2 Types of sequence alignment- pairwise and multiple alignment, local and Global alignment

2.3 Dot matrix comparison of sequences

2.4 Scoring matrices - PAM and BLOSUM

2.5 Pair wise sequence similarity search by BLAST and FASTA

2.6 Concepts of phylogeny- distance based (NJ Method) and Character based (ML method) ,Tree construction methods

Unit 3: Descriptive Biostatistics and Probability

3.1 Introduction to Biostatistics, kinds of data and variables, based on nature (numerical, discrete and continuous, categorical –ordinal and nominal), based on source (primary and secondary data) sample size, sampling methods and sampling errors

3.2 Data tabulation and representation methods, graphical methods(stem and leaf plot, line diagram, bar graphs, histogram, frequency polygon & frequency curve) diagrammaticmethod (pie diagram)

3.3 Measures of central tendency- arithmetic mean, median, mode (meritsand demerits)

3.4 Measures of dispersion- range, mean deviation, variance and standard deviation, Standard error and Co efficient of Variation -merits and demerits

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- 3.5 Concepts of probability-random experiment, events and Probability of an event, probability rules (addition and multiplication), uses of permutation and combinations, random variables
- 3.6 Probability distributions-Binomial, Poisson for discrete variables and Normal distribution for

## **Unit 4: Applications of Biostatistics**

- 4.1 Hypothesis testing- steps in testing for statistical hypothesis, null and alternative hypothesis level of significance-type 1 and type 2 errors
- 4.2 Test of significance- for small samples- student's t- test (one sample and two samples)
- 4.3 Test of significance- for large samples Z test for means and proportions
- 4.4 Chi-square test- and their applications -goodness of fit, test of independence
- 4.5 Analysis of Variance (ANOVA)- one way analysis
- 4.6 Correlation definition, simple linear analysis, Karl Pearson's correlation coefficient

#### **OPTIONAL I: PRACTICALS** BIOINFORMATICS AND BIOSTATISTICS

- Exploring web portals NCBI, EBI &ExPASy
- 2. Literature search through Pubmed and Pubmed Central
- Sequence retrieval from Genbank, ENA, Swissprot
- 4. Pairwise homology search by BLAST and FASTA
- Calculation of mean, median, mode, standard deviation, variance, standard error and coefficient of 5. variation
- Construction of bar diagram, pie diagram, line diagram, histogram
- Problems on hypothesis testing using Z- test. t-test and Chi-square test 7.
- 8. Problems on probability and probability distributions

#### Spotters

- Line diagram, bar diagram &pie diagrams 1.
- Histogram, frequency polygon & frequency curve 2.
- Normal Probable curve 3.
- 4. GenBank
- DDBJ 5.
- SWISS-PROT 6.
- 7. PROSITE
- 8. PIR
- BLAST
- 10. Pairwise alignment
- 11. Multiple sequence alignment
- 12. PAM and BLOSUM
- 13. Phylogenetic tree

## RECOMMENDED BOOKS

- 1. Khan &Khanum (2004), Fundamentals of Biostatistics, 11 Revised Edition, Ukaaz Publication
- 2. Bailey, N.T.J, Statistical methods in Biology, Cambridge Univ. Press
- 3. Fundamentals of Biostatistics, P HanmanthRao and K.Janardhan
- 4. Danial, W. W. Biostatistics, Wiley
- 5. Introduction to Bioinformatics by Aurther M lesk
- 6. Developing Bioinformatics Computer Skills by: Cynthia Gibas, Per Jambeck
- 7. Bioinformatics second edition by David M mount
- 8. Essential Bioinformatics by Jin Xiong
- 9. Bioinformatics Computing by Bryan Bergeron
- 10. Bioinformatics: Concepts, Skills & Applications by R.S. Rastogi
- 11. Queen, J. P., Quinn, G. P., & Keough, M. J. (2002). Experimental design and data analysis for biologists. Cambridge University Press

12. Mahajan, B.K. (2002). Methods in biostatistics. Jaypee Brothers Publishers

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B.Sc BIOTECHNOLOGY II YEAR (2021-22) SEMESTER- IV

SKILL ENHANCEMENT COURSE-3 (SEC-3) BS 401: MOLECULAR MARKERS IN PLANT BREEDING

## Unit 1: Molecular markers in Plant Breeding

- 1.1. Types of markers morphological, cytological, biochemical and genetic markers
- 1.2. Development of molecular markers scope in plant breeding; criteria for ideal molecular markers
- 1.3. Types of molecular markers
- 1.4. Hybridization based molecular markers RFLP
- 1.5. PCR based molecular markers RAPD, SSRs, AFLP
- 1.6. Sequence based molecular markers SNPs and DArTs

## Unit 2: Applications of Molecular markers in Plant Breeding

- 2.1. Segregating populations backcross, double haploid, F2&F3 families, RLLs
- 2.2. Linkage mapping and QTI, mapping
- 2.3. Marker Assisted Selection (MAS) procedure and applications
- 2.4. Map based cloning of genes
- 2.5. Fingerprinting fingerprinting genotypes; assessment of genetic similarity among genotypes; conservation, evaluation and use genetic resources
- 2.6. Hybrid testing

#### REFERENCE BOOKS

- 1. Gupta PK. 2010. Plant Biotechnology. Rastogi Publications.
- Chawla FIS. 2011. Introduction to Plant Biotechnology. Oxford and IBH Publishing 2. Co. PvtLtd.
- Chittaranjan K. 2006-07. Genome Mapping and Molecular Breeding in Plants. Vols. I-3. VU. Springer. 16
- Newbury HJ. 2003. Plant Molecular Breeding. Blackwell Publ. Weising K, Nybom H, 4. Wolff K & Kahl G. 2005. DNA Fingerprinting in Plants: Principles, Methods and Applications. Taylor & Francis.

## B.Sc BIOTECHNOLOGY III YEAR (2021-22)

#### SEMESTER- V GENERIC ELECTIVE (GE)

## BS 503: BASICS IN BIOTECHNOLOGY

## COURSE OUTCOMESCREDITS-4 TEACHING HOUR/WEEK-2

After completion of the course student will understand:

- About various tissue culture techniques to culture by invitro techniquesnand molecular farming
- Various fermentation techniques and process of production of the fermented foods and chemicals
- Applications of animal cell culture and use the assisted reproductive technology in livestock and its applications.
- Store and Retrieve drug related information using online tools and Comprehend the utility of tools & databases available in genomic & proteomics

### Unit: Agricultural Biotechnology

- 1.1. Plant tissue culture media, sterilization, culture types
- 1.2. Micro-propagation, Synthetic seeds, Somatic hybrids and haploid plants
- 1.3. Transgenic plants direct & indirect methods of gene transfer
- 1.4. Applications of transgenic plants improving productivity & nutritional quality
- 1.5. Applications of transgenic plants stress tolerant plants & molecular farming
- 1.6. Biofertilizers and biopesticides

## 3. Unit: Microbial and Industrial Biotechnology

- 2.1. Exploitation of micro-organisms and their products
- 2.2. Isolation, screening and selection of microorganisms for industrial products
- 2.3. Preservation of microorganisms
- 2.4. Strain development and improvement, strategies of strain improvement selection and recombination
- 2.5. Production of recombinant DNA vaccine, amino acids, vitamins
- 2.6. Single cell protein, dairy products, and penicillin and streptomycin production

## Unit: Animal and Medical Biotechnology

- 3.1. Cell culture technique and its applications
- 3.2. Animal breeding (selective breeding and cross breeding) and its limitations
- 3.3. In vitro techniques in animal improvement: in vitro fertilization & microinjection
- 3.4. Genetically modified animals: transgenic & knock-outs
- 3.5. Mouse models of disease: cancer and diabetes
- 3.6. Biotechniques: gel electrophoresis and PCR

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# Unit: Computer applications in Biotechnology

- 4.1. Scope of computer applications in Biotechnology
- 4.2. Biotechnology tools and resources role of the internet, free online tools, downloadable free
- 4.3. Biotechnology web portals NCBI, EBI, ExPASy
- 4.4. Biological databases: classification of databases the primary (Genbank), secondary (PIR) databases
- 4.5. Sequence databases DNA sequence databases (ENA &DDBJ)

4.6. Protein sequence databases (Swissprot& PROSITE)

#### B.Sc BIOTECHNOLOGY III YEAR (2021-22) SEMESTER- V

#### OPTIONAL- I (A) (DSE-1E)

## BS 504(A): PLANT BIOTECHNOLOGY

## COURSE OUTCOMESCREDITS-4 TEACHING HOUR/WEEK-4

After completion of the course student will understand:

- About various tissue culture techniques to culture by invitro techniques.
- Applications in plant tissue culture for Secondary metabolite production.
- The applications of genetic engineering like transgenic plants.
- Molecular farming for commercially synthesizing products such as vaccines, proteins, enzymes, etc.

#### 1 Unit: Fundamentals of Plant Tissue Culture

- 1.1. Introduction Plant tissue culture, totipotency of plant cells (dedifferentiation, re differentiation and regeneration)
- 1.2. Nutritional requirements for plant tissue culture: nutrient media macronutrients and micronutrients, media additives (carbon source, vitamins, amino acids); types of media
- 1.3. Plant growth regulators auxins, cytokinins and gibberilins
- 1.4. Preparation of media, sterilization, selection & surface sterilization of explant, inoculation, incubation and culture of plant tissue *in vitro*
- 1.5. Induction of callus cultures and cell suspension cultures
- 1.6. Organogenesis and somatic embryogenesis

## 2 Unit: Applications of Plant Tissue Culture

- 2.1. Meristem culture, micropropagation and their applications
- 2.2. Encapsulation and production of synthetics seeds and their applications
- 2.3. Cell suspension cultures (batch and continuous cultures) and applications
- 2.4. Protoplast isolation, culture and fusion development of somatic hybrids &cybrids and their applications
- 2.5. Somaclonal variation and its applications
- 2.6. Anther and pollen culture for production of haploids & their applications 2.7. Cryopreservation conservation of plant germplasm

## 3 Unit: Production of Transgenic Plants

- 3.1. Direct gene transfer techniques physical methods: microinjection, particle bombardment (gene gun) and electroporation & chemical methods
- 3.2. Molecular mechanism of Agrobucterium infection and features of Ti Plasmid

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- 3.3. Agrobacteirum mediated gene transfer using binary and co-integrate vectors
- 3.4. Viral vectors for gene transfer into plants
- 3.5. Selection of transgenic plants using reporter and selection marker genes
- 3.6. Genome editing -

## Unit: Applications of Transgenic Plants

- 4.1. Herbicide resistance in transgenic plants glyphosate tolerance
- 4.2. Insect resistant transgenic plants: Bt cotton, proteinase inhibitors, lectins
- 4.3. Virus, bacterial and fungal resistant transgenic plants
- 4.4. Abiotic Stress tolerance: drought, heat and salinity stress tolerant plants
- 4.5. Transgenic plants with enhanced nutritional value: vitamin A, oil, amino acids
- 4.6. Transgenic plants as bioreactors: edible vaccines, antibody production, biodegradable **Plastics**

#### OPTIONAL-I (A): PRACTICALS PLANT BIOTECHNOLOGY

- 1. Preparation of media for plant tissue culture
- 2. Sterilization methods of explants (seed, leaf, inter node & root) and inoculation
- 3. Establishment of callus cultures - from carrot/rice
- 4. Preparation of synthetic seeds
- 5. Meristem culture
- 6. Cell suspension cultures
- 7. Protoplast isolation and culture
- Agrobacterium mediated transformation 8.

#### Spotters

- Callus cultures 1.
- Sterilization techniques: autoclave and hot air Oven 2.
- Somatic embryos 3.
- Synthetic seeds 4.
- Meristem culture 5.
- Plant regeneration 6.
- Cell suspension cultures 7.
- Isolation of protoplasts 8.
- Particle bombardment (Gene gun) 9.
- Binary or co-integrate vectors 10.
- Gus gene expression in transgenic plant tissue 11.
- Golden Rice 12.

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## REFERENCE BOOKS

- Plant Tissue Culture and its Biotechnological Applications by W. Barz, E. Reinhard, M.H. 1. 2.
- Plant Tissue Culture by Akio Fujiwara 3.
- Frontiers of Plant Tissue Culture by Trevor A. Thorpe
- In vitro Haploid Production in Higher Plants by S. Mohan Jain, S.K. Sopory, R.E. 4. 5.
- Plant Tissue Culture: Theory and Practice by S.S. Bhojwani and A. Razdan 6.
- Plant Cell, Tissue and Organ Culture, Applied and Fundamental Aspects by Y.P.S. Bajaj and A. Reinhard

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## GOVERNMENT DEGREE COLLEGE FOR WOMEN, BEGUMPET, HYDERABAD.

## AUTONOMOUS (CBCS) B.Sc BIOTECHNOLOGY III YEAR (2021-22)

#### SEMESTER- VI

#### OPTIONAL PAPER I

## BS 601: IPR, BIOSAFETY AND ENTREPRENEURSHIP COURSE OUTCOMESCREDITS-4 TEACHING HOUR/WEEK-4

After completion of the course student will understand:

- Understanding, defining and differentiating different types of intellectual properties (IPs) and their roles in contributing to organizational competitiveness
- Exposing to the Legal management of IP and understanding of real life practice of IPM.
- Containment of potentially harmful biological agents and to reduce or eliminate exposure of laboratory workers, environment to potentially hazardous agents.
- The routes of exposure for a pathogen to a human being and demonstrate and assess the proper use of PPE, best practices.

#### 1. Unit: Intellectual Property rights

- 1.1. Intellectual Property meaning, nature
- 1.2. Significance and need of protection of intellectual property
- 1.3. Types of intellectual property rights: patent, trademarks, copyright, design registration, trade secret, geographical indicators, plant variety protection
- 1.4. Copyright: meaning, nature, historical evolution and significance
- 1.5. Ownership of copyright rights of authors and owners, trademarks
- 1.6. Plant varieties protection and plant breeding rights

#### 2. Unit: Patent laws

- 2.1. Patents concept of patent- historical overview of the patent law in India
- 2.2. Kinds of patents procedure for obtaining patent in India and in other countries
- 2.3. Patenting microbes and organisms- novelty, International Depository Authorities (IDAs), submitting details of the deposit
- 2.4. Patenting genes pros and cons, ethics, examples
- 2.5. Patenting markers and variants examples
- 2.6. Product vs process patent product life cycle and process design.

#### 3. Unit: Laboratory Management and Safety

- 3.1. Administration of laboratories, laboratory design, laboratory information management system
- 3.2. Laboratory safety good laboratory practice (GLP), biosafety levels
- 3.3. Basic principles of quality control (QC) and quality assurance (QA)
- 3.4. Handling of hazardous compounds chemicals, solvents, poisons, isotopes, explosives and biological strains



- 3.5. Storage of hazardous material
- 3.6. Disposal of biological and radioisotope wastes

## 4. Unit: Entrepreneurship

- 4.1. Concept, definition, structure and theories of entrepreneurship
- 4.2. Types of start-ups with examples
- 4.3. Types of entrepreneurship, environment, process of entrepreneurial development 4.4. Entrepreneurial culture, entrepreneurial leadership
- 4.5. Product planning and development project management, search for business idea, concept of projects, project identification

4.6. Promoting bio-entrepreneurship.

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## B.Sc BIOTECHNOLOGY III YEAR (2021-22)

## SEMESTER- VI

## OPTIONAL-II (A) (DSE-1F)

# BS 604(A): ANIMAL BIOTECHNOLOGY COURSE OUTCOMESCREDITS-4 TEACHING HOUR/WEEK-4

After completion of the course student will understand:

- To describe in vitro applications of animal cell culture
- To use the assisted reproductive technology practised in livestock and its applications
- To construct the techniques in production of cloned animal and its applications.
- To predict the ethical, social and moral issues related to cloning

## 1. Unit: Animal cell culture: principles and applications

- 1.1. Cell culture technique: cell culture media, sterilization techniques
- 1.2. Characteristic features of cell lines and cell line maintenance
- 1.3. Methods of isolation and separation of various cell types and establishment of cell lines
- 1.4. Properties and types of stem cells, culturing of embryonic stem cells and adult stem cells
- 1.5. Manipulation of cells: electroporation, transfection, transduction and microinjection
- 1.6. Applications of cell culture: manufacturing, toxicity testing and tissue engineering

#### 2. Unit: In vitro techniques in animal improvement

- 2.1. Principles of animal breeding: selective breeding, cross breeding and their limitations
- 2.2. Superovulation, collection of semen and ova
- 2.3. In vitro maturation of oocytes, artificial insemination
- 2.4. In vitro fertilization, embryo collection and embryo sexing
- 2.5. Somatic cell nuclear transfer, cloning of animals (example: Dolly)
- 2.6. Applications of in vitro techniques in animal improvement

#### 3. Unit: Molecular markers in animal genetics

- 3.1. Developments in livestock genomics
- 3.2. Molecular markers: types and characteristics
- 3.3. RFLP and RAPD
- 3.4. SNPs and their application in genotyping
- 3.5. Identification and isolation of desired genes of interest
- 3.6. Marker-assisted selection

## 4. Unit: Genetically modified organisms

- 4.1. Animal models and their significance in scientific research
- 4.2. Mouse models for cancer
- 4.3. Generation of transgenic mouse
- 4.4. Generation of gene knock-out mouse
- 4.5. Genetically modified mice as disease models

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4.6. Applications of genetically modified animals in understanding disease biology and drug development.

#### OPTIONAL-I (A): PRACTICALS ANIMAL BIOTECHNOLOGY

- 1. Preparation of animal cell culture media
- 2. Sterilization of cell culture media
- 3. Cell counting by microscopy
- 4. isolation of cells from chicken Liver
- 5. Establishment of primary cell culture: Liver/Spleen
- 6. Preparation of metaphase chromosomes
- 7. Culturing suspension cells
- 8. Culturing adherent cells

#### **Spotters**

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- 1. Microscope
- 2. CO2 incubator
- 3. Biosafety cabinet/ Laminar air flow
- 4. Trypan blue stained cells
- 5. Cell culture flasks and dishes
- 6. Metaphase slide
- 7. Autoclave
- 8. Centrifuge
- 9. Example of an RFLP
- 10. Microinjection into egg cells

#### REFERENCE BOOKS

- 1. Text book of Animal Biotechnology by B Singh. The Energy and Resources Institute (teri)
- 2. Genetics for Animal Sciences by WH Freeman. Van Vleck LD, Pollak EJ &Bltenacu EAB. 1987.
- 3. Cancer Cell Culture: Methods and Protocols: 731 (Methods in Molecular Biology) Humana; 2nd ed. 2011 edition (28 April 2011)
- 4. Genetic Engineering by V.K.Agarwal and P.S. Varma, S. Chand & Company Ltd, 2009

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